



CULTURE OF OYSTER MUSHROOM, *PLEUROTUS OSTREATUS* FROM AGRO WASTE – SUGARCANE MAGESY

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ABSTRACT :

Mushrooms are a group of macroscopic fungi and belonging to the class Basidiomycetes. About 2500 varieties of mushrooms are available in the world out of which only 200 species are edible. The oyster mushrooms are highly edible and rich in nutrients such as proteins and vitamins. Mushrooms are cholesterol free diet and good for diabetic patients. Sugarcane magesy is a solid agriculture waste and it contains sucrose and act as excellent substratum for mushroom growth. The present study aimed to discussion oyster mushroom culture and solid waste management of sugarcane magesy.

KEYWORDS : Agricultural waste, sugarcane magesy, *Pleurotus ostreatus* production.

INTRODUCTION

Oyster mushroom (*Pleurotus ostreatus*) is an edible mushroom. It contains adequate amount of phosphorous, iron, protein, lipid, riboflavin and thiamine. Oyster mushroom fresh fruiting bodies indicates a high quantity of moisture (90.8%), where as dry as well as fresh oyster mushrooms are rich in carbohydrate (57.6%), protein (30.4%), fiber (8.7%), fat (2.2%) and ash (9.8%) with 345 kilocalories energy value on 100 g dry weight. Mushrooms are an excellent source of minerals and protein and also known as the vegetarian's meat [44]. The proteins of mushroom are considered to be intermediate between that of vegetables and animals [42]. The amino acids essential for human body are present in oyster mushroom [36]. *P. ostreatus* productivity is maximum in a short time providing more protein per unit area than any other crop [35]. The high content of nitrogen and protein in oyster mushroom increases the biological efficiency and yield while it reduces the growth period. *P. ostreatus* contains 18 essential amino acids such as methionine, isoleucine, lysine, glutamic acid, cysteine, aspartic acid phenylalanine, tyrosine, tryptophan, valine, arginine, histidine, alanine, glycine serine and proline [26]. The vitamins such as niacin, riboflavin and thiamin are found in oyster mushroom. The minerals such as ferrous sulfate, phosphorus, sodium and calcium are present in oyster mushroom [60]. It is used for both medicine and food [6]. According to Furlani and Godoy [32], mushrooms are considered as food with high nutritional value and delicious taste. Due to their low caloric



value they are suitable for diets. The *P. ostreatus* produces metabolites of medicinal and pharmacological interest, such as antimicrobials, immunostimulants, antioxidants and antitumourals [28, 56, 38]. The substrate source, spawn quality, strain and compost affect the performance and growth of oyster mushroom [77, 39]. The oyster mushrooms are known for anti-inflammatory and immune-modulator effects [48]. The cultivation is easy under both temperate and tropical

climatic conditions and they are cultivated and harvested all over the year [7].

The present research was carried out to evaluate different agricultural substrates for growth and production of *Pleurotus ostreatus*, and to check quality of mushroom grown on sugarcane magesy. The objectives of this research are to evaluate paddy straw substrate for the growth and production of oyster mushroom and to find the effect of substrate on the quality of oyster mushroom.

MATERIALS AND METHODS

The culture was developed from the fruiting bodies growing at the Mushroom House. Malt Extract Agar (MEA) was employed as medium for the growth of pure culture of *Pleurotus ostreatus*. For one liter of the culture medium prepared, 20g Malt, 20g Dextrose, 20g Agar, 1g Peptone and distilled water were used to make the volume one liter. After autoclaving at 15 psi for 15 minutes, the pH of the medium was kept 6.5. Streptomycin was added in the sterilized medium @1g/ L when the medium was cooled to about 40°C to stop the bacterial contamination. The inoculation was made by cutting small pieces of the mushroom tissues from the cap to downwards with a sterile blade inside the laminar flow unit. A small piece of fruiting body was taken and inserted in to petridish. Plates were sealed with Parafilm and incubated at 25°C. After 3 to 4 days, the white mycelium has covered the agar surface. When the mycelium fully covered the plates, then they are kept in refrigerator for the preparation of spawn. The prepared cultures were multiplied and maintained through standard methods.

PREPARATION OF SPAWN

Spawn was prepared in jam bottles. The method of Jain (2005) was followed with some modifications. The paddy grains were boiled for 20-30 min. After boiling, excess water was drained off by spreading the grains on a wire mesh. Chalk powder (calcium carbonate) and gypsum (calcium sulphate) were added to the grains @ of 2% and 0.5%, respectively on dry weight basis. The jam bottles containing paddy grains were sterilized in an autoclave for one hour at 121°C. The grains were allowed to cool at room temperature and inoculated with actively growing mycelium of *Pleurotus ostreatus* from malt extract slants inside the laminar flow unit and incubated at (27 ± 2°C) for mycelial growth without any light for 10-15 days until the mycelium fully covered the grains. Paddy straw substrate was sundried and broken into small pieces and soaked in water to maintain 65-75% moisture content. The soaked substrate was piled up after and covered with polythene sheet. Substrate was allowed to ferment for hours and spread on floor for evaporation of excess moisture. The experiment was laid by using Completely Randomized design having ten replications.

STERILIZATION AND INCUBATION

The sugarcane magesy substrate was filled in 30 glass bags (12×18 inch) and bag mouth was loosely tied with fiber thread. Bags were sterilized in an air tight container at 15 psi for one hour. The sterilized bags cooled for 2-3 hours were inoculated with spawn @ 25 gm per kg bag [21]. The sterilized bags were then kept in mushroom house at ambient temperature. These bags after inoculation were then incubated for spawn running in a mushroom house under darkness at ambient temperature.

CROPPING

When mycelia fully covered the substrates after 15-16 days, the bags were torn apart to open the substrates. The compact substrates were irrigated at least twice a day by sprinkling fresh water. After 7–8 days of the opening of bags small size pin heads (4-5 cm in diameter) appeared on all sides of the bags. These pinheads attained the full size in about 2-3 days and when fruiting body fully matured then they were harvested. The pin heads appearance time was also recorded. The oxygen needed for the fruiting bodies development was fulfilled by running fan several times daily. The experiment was laid out in a complete randomized design (CRD) having 5 treatments and ten replications. Data were recorded on the following parameters. Data on spawn running was recorded in days on substrate.

APPEARANCE AND MATURATION OF PINHEAD

After the completion of spawn running the pinheads appearance of *Pleurotus ostreatus* was observed. The data was recorded in days taken from spawning to the appearance of pinheads in of the substrate. When the pinheads reached to maximum size then time period was recorded in days from appearance of pinheads to maturation of pinhead treatment.

YIELD

The data on the weight of mushroom in gram was recorded for the harvesting of mushroom. The first and respective harvesting done at maturity of fruiting bodies was noted. The total yield of basidiocarp was measured.

PHYSICAL, CHEMICAL AND BIOCHEMICAL DETERMINATION

Mushroom sample used in the study was oven dried at 105 ± 5 °C for 24 hours till constant weight was achieved. Moisture, ash content, crude fiber content, protein and crude fat contents were tested according the procedure followed by Raghuramulu *et al.*, 2003. Vitamin C, Phosphorous, Iron, Zinc and Nitrogen free extract content were estimated according standard procedure.

RESULTS

The result shown in Table 1 indicates that fastest spawn running took place on sugarcane magesy and it takes 29 days. Result of the experiment about the appearance of pinheads and their maturity are presented in Table 1. It was observed that time taken for first appearance of pinhead after spawning of the substrates was fastest (40 days) in sugarcane magesy.

The sugarcane magesy took 40 days from appearance of pinheads to the maturity of pinheads. In response to maturity of pinheads, the statistical analysis shows that paddy straw is highly significant. The average yield of the substrate from first flush was 480 gram, respectively. The total yield was determined by taking the weight of mushroom obtained after 1st, 2nd and 3rd flush. The moisture content of mature fruiting bodies of *Pleurotus ostreatus* cultivated on paddy straw is shown in Table 2. The Table indicates that higher moisture content (91%) was observed. The total Ash content of mature fruiting bodies of *P. ostreatus* grown on different agricultural waste paddy straw is shown in table 2.

The protein content of mature fruiting bodies of *P. ostreatus* cultivated on paddy straw and the result indicates that high protein content 9.6 gram level. From the Table it was observed that the paddy straw have the medium crude fat and crude fiber content 4 % and 2.50 % level. The vitamin C contents of mature fruiting bodies of *P. ostreatus* cultivated on paddy straw is presented in table 3. The result indicates that paddy straw exhibit vitamin C content of 10.47mg/ 100gram. From the result it was evident that paddy straw has phosphorous content of 0.33 mg/100 gram. Iron content was observed that paddy straw have 0.009 mg/ 100 gram. The zinc and nitrogen free extract of mature fruiting bodies of *P. ostreatus* cultivated on paddy straw was observed that 0.006 and 2.31 gram/100 gram.

DISCUSSION

The study indicated that *Pleurotus ostreatus* can be successfully grown on almost all agricultural wastes but paddy straw gave better results. The structure of substrates is also important because it help in the penetration of mycelium. Paddy spawn is commonly used for mushroom cultivation because the growth of mycelium is faster on other grains. Present study revealed that the growth rate of mycelium is faster. Paddy straw has been used for the cultivation of oyster mushroom since the beginning of 19th century and it has been cultivated in many countries under natural condition [69].

Sugarcane magesy is the natural substrate on which oyster mushrooms are cultivated leading to name the mushroom as delicious straw reported by Fasidi (1996) [31]. According to Balasubramanya and Kathe (1996) [8], the fungus *Penicillium* spp. and *Trichoderma* spp competed with *Pleurotus* spp. after pasteurisation with hot water (80°C for 2h) probably due to the partial breakdown of hemicellulose,

cellulose and thus making them available to competitors. The successful mushroom cultivation depends on the purity and quality of spawn. Nita Bahl (1984) [58] reported that grain spawn is now almost universally used. The difference in days for full mycelial running on different substrates might be due to variation in their chemical composition and C: N ratio as reported by Bhatti et al. (1987) [11]. The results recorded on spawn running on different substrates were almost similar to the findings of Shah et al. (2004). They reported that the spawn running took 16-25 days after inoculation. Similar results were also reported by Tan (1981) [90] who reported 21 days for complete spawn running on cotton waste. Similar findings were also reported by Jiskani et al. (1999) [40]. The pinheads appeared earlier in paddy straw than other substrates.

Khan et al. (1981) [44] observed the appearance of 30 pin-heads of *P. ostreatus* (Strain, -467) in 36 days, the same of *P. sajor-caju* and *P. ostreatus* in 40 and 46 days, after spawning. Tan (1981) [90] recorded 23-26 days for the appearance of pinheads. Ramzan (1982) [73] observed 20-40 days of five *P. ostreatus* strains on wheat and paddy straw. Patra and Pani (1995) [64] recorded 20-24 days on paddy straw.

Khan et al. (1981) [44] recorded 21-28 days for the maturity of pinheads in case of cotton boll locules. Khanna and Garcha (1981) [45] recorded 20-24 days for the maturity of pinheads on paddy straw and Tan (1981) [90] observed a month for the maturity of pinheads on cotton waste. There is difference in yield obtained from different substrates. The present finding indicates that highest yield was obtained in wheat straw followed by paddy straw. Bhatti et al. 1987 [13] observed the highest yields with the shortest incubation period in case of wheat straw. It was generalized from the data that first flush yield was highest in all treatments followed by second and third flush. Other scientists also recorded similar results. Zadrazil (1973) [95] got 2 flushes, Tan (1981) [90] got three flushes, Ramzan (1982) [73] obtained 3-5 flushes from wheat and paddy straw and Bhatti (1984) [12] got 4-6 flushes from different substrates. Kausar and Iqbal (1994) [43] investigated that B.E varied from 18.6 to 83.5% on the basis of different nitrogen supplements amended with straw. Jiskani et al., (1999) [40] obtained 24 and 7.6% fresh and dry yield on the basis of substrate dry weight by using wheat straw. Jiskani (1999) [40] reported that one kg of dry substrate can produce one kg of fresh mushroom which is the 100% substrate dry weight. According to Bughio (2001) [16] the maximum dry and fresh (wet) yield percentage on substrate dry weight basis (29.61 to 77.91 and 5.91 to 21.70) were obtained from wheat straw using in combination with sugarcane bagasse, paddy straw, cotton boll locules and sorghum leaves.

Graham and Clyde (1985) [33] recorded 80-120 percent biological efficiency of *P. sajor-caju* on cotton waste. Moonmoon et al., (2010) [55] studied *P. eryngii* King Oyster mushroom on paddy straw and saw dust in Bangladesh and found that saw dust. Nunez and Mendoza (2002) [59] reported that the biological efficiency of the studied substrates varied from 106.2 to 50.8% of *Pleurotus flabellatus*. The moisture content of the studied mushroom ranged from 88.20% to 93.44%. Moisture percentage in mushroom depends on the maturity of fruiting bodies, species and storage conditions during packaging or processing [34]. Present study revealed that the ash content of studied mushroom ranged between 0.33% - 1.006%. The amount of ash was higher in sorghum than other substrates. The amount of ash depends on salt content in substrates [68]. (Bonatti et al. (2004) [15] reported 0.5-0.6% of ash in dried *P. sajor* whereas Alam et al. (2008) [5] recorded 1.1 and 8.28 g/100g in fresh and dried *P. sajor-caju*, respectively. El-Kattan et al., (1991) [27] reported 8.00% and 6.60% ash content of *P. ostreatus* on soybean and paddy straw, respectively. The analysis of mushroom composition indicated that sugarcane bagasse gave high amount of proteins and amino acids from other substrates.

The protein content usually ranges between 20–30% on a dry weight basis. The nitrogen present in substrate after spawn running enhances the mushroom yield and quality, in addition it help in bioconversion and bioaccumulation efficiency [63]. The fat content of *P. bellatus* was 10% grown on maize straw being the highest followed by sorghum straw (2.50%). The percent content were similar as reported in earlier studies [63, 62]. The fat content ranged between 2.56% to 2.82% on dry weight basis. The fat content of dried *P. sajor-caju* was 5.26 and 4.99% on paddy straw and banana straw, respectively [15]. The highest crude fiber (3.5%) was obtained from mushroom on wheat straw followed by paddy straw (2.75%). Other agro wastes also yielded appreciable level of crude fiber. These results were confirmed with the findings of Singh et al.

(2003) [83], Bonatti et al. (2004) [15], Kadlag et al., (1998) [41] and Sharma & Madan (1993) [81]. The amount of protein of *P. bellatus* found in this study is near about similar to the results of Rai and Sohi (1988) [71] and Alam et al. (2007) [4]. But fiber and ash content are different from the report of Rai and Sohi (1988) [71], however relevant to Alam et al. (2007) [4].

Protein content in *P. ostreatus* were also similar to the findings of Banik and Nandi (2004) [9] as well as fat value of *P. ostreatus* is relevant to the findings of Shashirekha et al. (2005) [82]. The results indicated that not only the protein content of the substrate but also nature of protein in the substrate influences the protein content of the fruiting bodies [92]. The amount of vitamins C were maximum (11.35mg/100 gram) in paddy, maize and sorghum straw. Bano (1976) recorded 13.0 to 14.70 mg/100 g ascorbic acid in various mushroom species. Present study revealed that sugarcane bagasse have high phosphorous amount as compare to other substrates. Similar results were observed by Chang et al. (1981) [20] and Alam et al. (2007) [4], but differ from [79] who recorded 0.97% phosphorus in oyster mushrooms grown on sawdust. The amount of phosphorous was maximum on soybean straw (920mg/100gm) whereas least was found on soybean straw and wheat straw (800 mg/100gm) [17]. Phosphorus content of *P. bellatus* ranged from 790 – 1000 mg/100g [65].

CONCLUSION

Sugarcane magesy was found most suitable substrate for mushroom cultivation. The spawn running, appearance and maturity of pinheads were fastest and also showed the highest flush wise yield, total yield and biological efficiency. Moisture, ash, nitrogen free extract and vitamin C were maximum in paddy straw. The highest protein content was obtained from paddy straw. Meanwhile the percentage of crude fat and iron were highest. Farmers are advised to use wheat straw for *Pleurotus ostreatus* cultivation for bumpy production. However as more nutrients have been found in mushroom grown on paddy straw, so they can also use sugarcane magesy as substrate for the production of high quality mushrooms.

Table 1 Production of *Pleurotus ostreatus* on sugarcane magesy substrate

S.No	Paramerters	Days
1	Days to spawn	29
2	Days of pin head appearance	40
3	Days to maturity	42
4	Yield (gm)	480

Number of replications n=8

Table 2 Percent moisture, ash, protein, crude fat and crude fiber in oyster mushroom (*Pleurotus ostreatus*) grown on sugarcane magesy substrate

S.No	Paramerters	Percentage (%)
1	Moisture	91
2	Ash	0.33
3	Protein	9.6
4	Crude fat	6
5	Crude fiber	2.55

Number of replications n=8

Table 3 Vitamin C, Phosphorous, Iron, Zinc and Nitrogen free extract in *Pleurotus ostreatus* (mg/100 gram) grown on sugarcane magesy substrate

S.No	Paramerters	mg/gram
1	Vitamin C	10.47
2	Phosphorous	0.33
3	Iron	0.006
4	Zinc	0.006
5	Nitrogen Free Extract	2.31

Number of replications n=8

REFERENCES

1. Adamovic M, Grubic G, Milenkovic I, Jovanovic R, Protic R, Sretenovic L, Stoicevic LJ. The biodegradation of wheat straw by *Pleurotus ostreatus* mushrooms and its use in cattle feeding. *Animal Feed Science and Technology*, 1998; 71:357-362.
2. Adebayo GJB, Omolara BN, Toyin AE. Evaluation of yield of oyster mushroom (*Pleurotus pulmonarius*) grown on cotton waste and cassava peel. *African Journal of Biotechnology*. 2009; 8:215-218.
3. Adejoye OD, Adebayo Tayo BC, Ogunjobi AA, Olaoye OA, Fadahunsi FI. Effect of carbon, nitrogen and mineral sources on growth of *Pleurotus florida*, Nigeria edible mushroom. *African Journal of Biotechnology*. 2006; 5:1355-1359.
4. Alam N, Khan A, Hossain MS, Amin SMR, Khan LA. Nutritional analysis of dietary Mushroom- *Pleurotus florida* Eger and *Pleurotus sajor-caju* (Fr.) Singer. *Bangladesh Journal of Mushroom*. 2007; 1:1-7.
5. Alam N, Amin R, Khan A, Ara I, Shim MJ, Lee MW et al. Nutritional analysis of cultivated mushrooms in Bangladesh *Pleurotus ostreatus*, *Pleurotus sajor-caju*, *Pleurotus florida* and *Calocybe indica*. *Mycobiology*, 2008; 36:228-232.
6. Alice B, Kustudia M. Mushroom cultivation and marketing. NCAT ATTRA Publication No. IP087, 2004; 30:25-29.
7. Amin SMR, Nirod CS, Moonmoon M, Khandaker J, Rahman M. Officer's training manual. national Mushroom development and extension Centre, Savar, Dhaka, Bangladesh, 2007, 7-17.
8. Balasubramanya RH, Kathe AA. An inexpensive pretreatment of cellulosic materials for growing edible oyster mushrooms. *Biol. Resour. Technol*, 1996; 57:303305.
9. Banik KS, Nandi R. Effect of supplementation of paddy straw with bagass residual slurry manure on the yield, protein and mineral contents of oyster mushroom. *Indian Crop Production*, 2004; 20:311-319.
10. Bano Z. The nutritive values of mushrooms. In: Proceeding of the first symposium on survey and cultivation of edible mushrooms in India. R. R. L. Shrinagar. 1976; 2:172.
11. Bhatti A, Kustudia M. Mushroom cultivation and marketing. In: Horticulture Production Guide, NCAT, 2004. <http://www.attra.org/attra-pub/PDF/mushroom.pdf> 24p
12. Bhatti MA. Mushrooms as commercial crop. *Progressive Farming*, 1984; 4:5-10.
13. Bhatti MA, Mir FA, Siddiq M. Effect of different bedding materials on relative yield of oyster mushroom in the successive flushes. *Pakistan Journal of Agriculture Research*. 1987; 8:256-259.
14. Bilal AW, Bodha RH, Wani AH. Nutritional and medicinal importance of mushrooms. *J. Medicinal Plants Research*. 2010; 4:2598-2604.
15. Bonatti M, Karnopp P, Soares HM, Furlan SA. Evaluation of *Pleurotus ostreatus* and *Pleurotus sajorcaju* nutritional characteristics when cultivated in different lignocellulosic waste. *Food Chemistry*, 2004; 8:425-428.
16. Bughio I. Yield performance of oyster mushroom, *Pleurotus ostreatus* (Jacq. ex Fr.) Kummer on combination of different straws. M. Sc. Thesis, Deptt. of P. Path. S.A.U. Tandojam, 2001, 69.
17. Caglarırmak N. The nutrients of exotic mushrooms (*Lentinula edodes* and *Pleurotus* species) and an estimated approach to the volatile compounds. *Food Chemistry*, 2007; 105:1188-1194.
18. Chang ST, Miles PG. *Edible mushrooms and their cultivation*, Florida: CRC press, 1989; 30:25-29.
19. Chang ST. World production of cultivated edible and medicinal mushrooms in 1997 with emphasis on *Lentinus edodes*. China. *International J. Medicinal Mushrooms*. 1999; 1:291-300.
20. Chang ST, Lau OW, Cho KY. The cultivation and nutritional value of *Pleurotus sajor-caju*. *European Journal of Applied Microbiology and Biotechnology*. 1981; 12:58-62.
21. Choi KW. Oyster mushroom cultivation: shelf or bag, 2003. <http://www.mushworld.com>
22. Cohen A, Dorffling K, Bettin D, Hahn H. Abscisic acid and cytokinins as possible root to shoot signals in xylem sap of paddy plants in drying soil. *Australian Journal of Plant Physiology*. 2002; 20:109-115.
23. Courvoisier M. Les champignons comestibles dans le monde. *Bul. Fed. Nat. Syn. Champ*, 1999; 82:829-837.

24. Diana F, Indrea D, Apahidean AS, Apahidean M, Pop R, Moldovan Z, Măniu Ńiu D, Ganea R, Paven I. Importance of substrat disinfection on Oyster mushroom (*Pleurotus* sp.) culture. Not. Bot. Hort. Agrobot. Cluj, 2006; 34:1-6.
25. Dias ES, Koshikumo EMS, Schwan RF, Silva F. Cultivo do cogumelo *Pleurotus sajor-caju* em diferentes resíduos agrícolas. Ciênc. Agrotechnology, 2003; 27:1363-1369.
26. Djarijah NM, Djarijah AB. Budidaya jamur tiram. penerbit kanisius, Yogyakarta. Inheritance of some marker genes in *Setaria italica*. Theor. Appl. Genet, 2001; 71:57-60.
27. El – Kattan MH, Helmy ZA, Abdel H, Leithy Mel, Abdelkawi KA. Studies on cultivation techniques and Chemical composition of Oyster mushrooms. Mushroom Journal for tropics. 1991; 11:59-66.
28. Elmastas M, Isildak O, Turkekel I, Temur N. Determination of antioxidant activity and anti-oxidant compounds in wild edible mushrooms. J. Food Compos Anal. 2007; 20:337-345.
29. Erkel I. Effects of different growing medium on yield of *Pleurotus ostreatus* and *Pleurotus florida* cultivation, Fourth congress of edible mushroom of Turkey, Yalova, 1992; 1:56-60.
30. Fanadzo M, Zireva DT, Dube E, Mashingaidze AB. Evaluation of various substrates and complements for biological efficiency of *Pleurotus sajor-caju* and *Pleurotus ostreatus*. African Journal of Biotechnology. 2010; 9:2756-2761.
31. Fasidi IO. Studies on oyster mushroom (*Pleurotus florida*) cultivation on agricultural wastes and proximate composition of stored mushrooms. Food chem, 1996; 55:161-163.
32. Furlani RPZ, Godoy HT. Valor nutricional decogumelos comestíveis. Scientific. Technol. Alim, 2007; 27:154-157.
33. Graham, Clyde M. *Pleurotus* mushroom kits. Mush. Newsletter for the tropics, 1985; 6(2):10.
34. Guillamón E, García Lafuente A, Lozano M, Arrigo MD, Rostagno MA, Villares A et al. Edible mushrooms: role in the prevention of cardiovascular diseases. Fitoterapia, 2010; 81:715-772.
35. Gupta VK Prasad, Bakshi KS, Langar MPS. Improving nutritive value of roundnut shells through fungal cultivation. Agric. Wastes, 1986; 16:161-169.
36. Hayes WA, Haddad SP. The nutritive value of mushrooms. Mushroom. J. 1976; 30:204. activities of shiitake (*Lentinula edodes*) extracts obtained by organic solvents and supercritical fluids. Journal of Food England. 80:631-638.
37. Holker U, Holker M, Lenz J. Biotechnological advantages of laboratory scale solid state fermentation with fungi. Applied Microbiology and Biotechnology, 2004; 64:175-186.
38. Israilides C, Kletsas D, Arapoglou A, Philippoussis H, Pratsinis H et al. In vitro cytostatic and immunomodulatory properties of the medicinal mushroom *Lentinula edodes*. Phytomedicine, 2008; 15:512-519.
39. Jafarpour M, Zand AJ, Dehdashtizadeh B, Egh Sh. Evaluation of agricultural wastes and food complements usage on growth characteristics of *Pleurotus ostreatus*. African Journal of Agriculture Research. 2010; 5:32913295.
40. Jiskani MM, Pathan MA, Wagan KH. Yield performance of oyster mushroom *Pleurotus florida* (strain PK, 401) on different substrates. Pakistan Journal of Agriculture Engineering and Veterinary Sciences. 1999; 15:26-29.
41. Kadlag GK, Wani PV, Sawant DM. Comparative performance of different *Pleurotus* spp on wheat and green gram straw. Maharashtra Agric. Univ, 1998; 23:2586.
42. Kurtzman RHJ. Summary of mushroom culture. In Proceedings of Seminar of Mushroom Research and Production PARC, Karachi–Pakistan, 1975, 15-22.
43. Kausar T, Iqbal SH. Supplementation of paddy straw with various nitrogen sources to improve the yield of *P. sajorcaju*. Pak. J. Sci Ind Res. 1994; 37:615-619.
44. Khan SM, Kausar AG, Ali MA. Yield performance of different strains of oyster mushroom (*Pleurotus* spp.) on paddy straw in Pakistan. Mush. Sci, 1981; 11:675-678.
45. Khanna P, Garcha HC. Introducing the cultivation of *Pleurotus florida* in the plains of India. Mush. Sci, 1981; 11:655-665.

46. Kikuchi M, Tamakawa K, Hiroshimo K, Aihara Y, Mishimu V, Seki T. Survey contents of metals in edible mushrooms. *Journal of Hygienic Society of Japan*. 1984; 25:534-535.
47. Kwon H, Kim BS. *Mushroom Growers' Handbook: Shiitake Cultivation*, Mushroomworld, Korea. 2004, 260.
48. Lavi I, Levinson D, Peri I, Hadar Y, Schwartz B. Orally administered glucans from the edible mushroom (*Pleurotus pulmonarius*) reduce acute inflammation in dextran sulfatesodium induced experimental colitis. *British Journal of Nutrition*. 2010; 103:393-402.
49. Mandhare VK. Productivity of *Pleurotus* sp on different substrates and its effect on Nutritional Indices of spent straw. Ph.D. Thesis. Marathwada Agricultural Univ. Parbhani. 2000.
50. Mane VP, Patil SS, Syed AA, MMV. Baig Bioconversion of low quality lignocellulosic agricultural waste into edible protein by *Pleurotus sajor-caju* (Fr.) Singer. *J Zhejiang Univ Sci B*. 2007; 8:745-751.
51. Manzi P, Gambelli L, Marconi S, Vivanti V, Pizzoferrato L. Nutrients in edible mushrooms an inter species comparative study. *Food Chemistry*, 65: 477-482.
52. Martínez, Carrera D. *Mushroom*. McGraw-Hill Encyclopedia of Science and Technology. McGraw Hill Inc New York, 2002; 7:91-96.
53. Mehta V, Gupta JK, Kaushal S. Cultivation of *Pleurotus florida* mushroom on paddy straw and biogas production from the spent straw. *World Journal of Microbiology and Biotechnology*. 1990; 6:366-370.
54. Miroslawa Z. Influence of substrate pasteurisation methods on the yielding of some *Pleurotus* cultivars. *Res InstVegetable Crops*, 1991; 51:35-40.
55. Moonmoon MM, Uddin N, Ahmed S, Shelly NJ, Khan MA. Cultivation of different strains of king oyster mushroom (*Pleurotus eryngii*) on saw dust and paddy straw in Bangladesh. *Saudi J. of Bio Sci*. 2010; 17:341-345.
56. Moradali F, Mostafavi H, Ghods S, Hedjaroude A. Immunomodulating and anticancer agents in the realm of macromycetes fungi (macrofungi). *Int. Immunopharmacol*, 2007; 7:701-724.
57. Nataraja S, Danda Sanjeev Pradeep Y, Ponmurugan P, Kannan N. Effect of different substrates on mushroom cultivation under Namakkal district climate. National level biological conference on biotechnology-A Boon humanity, (abstract). Muthayammal college of Arts and science, Rasipuram, 2005, 7.
58. Nita B. *Hand book on mushroom*. Oxford and IBH publ. Co., New Delhi, Bombay, Calcutta, 1984, 52-53.
59. Nunez JP, Mendoza CG. Submerged fermentation of lignocellulosic wastes under moderate temperature conditions for oyster mushroom growing substrates. *Mushroom Biology and Mushroom Products*, 2002; 5:545-549.
60. Pandey RS, Ghosh SK. *A Handbook on cultivation of mushroom*. Emkay Publications Delhi, 1996; 3:134-140.
61. Pant D, Reddy UG, Adholeya A. Cultivation of oyster mushrooms on wheat straw and bagass substrate amended with distillery effluent. *World Journal of Microbiology and Biotechnology*. 2006; 22:267-275.
62. Patil SS, Dakore HG. Comparative study on yield performance and Nutritive value of oyster mushroom on soybean straw. *Bioinfolet*, 2007; 4:57-59.
63. Patill SS, Kadam RM, Shinde SL, Deshmukh SA. Effect of different substrate on productivity and proximate composition of *P. florida*. *International journal of plant sciences*. 2008; 3:151-153.
64. Patra AK, Pani BK. Yield response of different species of oyster mushroom (*Pleurotus*) to paddy straw. *Current Agric. Res*, 1995; 8:11-14.
65. Patrabansh S, Madan M. Studies on cultivation, biological efficiency and chemical analysis of *Pleurotus sajor caju* (Fr.) Singer on different bio wastes. *Acta Biotechnologica*, 1997; 17:107-122.
66. Peksen A, Yakupoglu G. Tea waste as a complement for the cultivation of *Ganoderma lucidum*. *World Journal of Microbiology and Biotechnology*. 2009; 25:611-618.
67. Peter, Oei. *Mushroom Cultivation*. 3rd Edition. Appropriate technology for mushroom growers. Backhuys Publishers, Leiden. The Netherlands, 2003; 5:782-787.
68. Pomeranz Y, Meloan CE. *Food Analysis: Theory and practice*. United States of America. Aspen *Pleurotus florida* (Strain Pk-401) on different substrates. *Pak. Jr. Agri., Agril. Engg. Vet. Sci*, 2000; 15:26-29.

69. Quimio TH, Chang ST, Royse DJ. Technical Guidelines for Mushroom Growing in the Tropics, FAO, Plant Production and Protection paper No 106. Rome, Italy. 1990, 154.
70. Raghuramulu N, Madhavan NK, Kalyanasundaram S. National Institute of Nutrition. Indian Council of Medical Research, Hyderabad, India. Manual of Laboratory Techniques, 2003, 56-58.
71. Rai RD, Sohi HS. How protein are rich in mushrooms. Indian Horticulture. 33: 2-3. Rajarathnam, S., Z. Bano.1989. Biotransformations of natural lignocellulosic wastes: commercial applications and implications, Critical Reviews in Food Science and Nutrition, 1988; 28:31-113.
72. Rajarathnam S, Bano Z. Biotransformations of natural lignocellulosic wastes: commercial applications and implications, Critical Reviews in Food Science and Nutrition, 1989; 28:31-113.
73. Ramzan M. Studies on the cultivation of oyster mushroom (*Pleurotus* spp.) in Faisalabad. M.Sc. Thesis, Department P. Pathology, Faculty of Agriculture, University of Agriculture, Faisalabad, 1982.
74. Rathore VRS, Thakore BBL. Effect of different substrates on the production and nutritional value of sporophores of *Pleurotus florida* (Eger) Nom. Nud. Journal of Mycology and Plant Pathology, 2004; 34:6668.
75. Royse DJ. Effects of spawn run time and substrate nutrition on yield and size of the shiitake mushroom. Mycologia, 1985; 75:756-762.
76. Royse DJ. Influence of spawn rate and commercial delayed release of nutrient levels on *Pleurotus conocopiae* yield, size and time to production, Applied Microbiology and Biotechnology, 2002; 17:191-200.
77. Royse DJ, Fales SL, Karunanandaa K. Influence of formaldehyde-treated soybean and commercial nutrient complementation on mushroom (*Pleurotus sajor-caju*) yield and in-vitro dry matter digestibility of spent substrate. Appl. Microbiol. Biotechnol, 2004; 36:425429.
78. Sanchez C. Cultivation of *Pleurotus ostreatus* and other edible mushrooms. Appl. Microbiol. Biotechnol, 2010; 85:1321-1337.
79. Sarker NC, Hossain MM, Sultana N, Mian IH, Karim AJMS, Amin SMR. Effect of different levels of pH on the growth and yield of *Pleurotus ostreatus* (Jacquin ex. Fr.) Kummer. Bangladesh J. Mush. 2007; 1:57-62.
80. Schmidt O. Experiments with mushroom cultivation on wood waste. Plant Research and Development, 1986; 24:85-92.
81. Sharma S, Madan M. Microbial protein from leguminous and non-leguminous substrates. Acta Biotechnologica, 1993; 13:131-139.
82. Shashirekha M, Rajarathnam N, Bano SZ. Effects of supplementing paddy straw growth substrate with cotton seeds on the analytical characteristics of the mushroom, *Pleurotus florida* (Block & Tsao). Food Chem, 2005; 92:255-259.
83. Singh NI, Singh TC, Devi MB. Nutritional composition, processing and preservation of the edible mushroom found in Manipur for sustainable economic development. J. Mycological research. 2003; 41:243-244.
84. Sivaprakasam K, Kandaswamy TK. Waste materials for the cultivation of *P. sajor-caju*. Mushroom J. 1981; 101:178-179.
85. Stamets P. The role of mushrooms in nature. Growing gourmet and medicinal mushrooms, Ten speed press, Berkeley, California, USA, 2000, 10.
86. Stamets P. Mycelium Running: How Mushroom Can Help Save the World, p: 574. Ten Speed Press, Berkeley and Toronto, 2005.
87. Stanley HO, Awi-Waadu GD. Effect of substrates of spawn production on mycelial growth of Oyster mushroom species. Research Journal of Applied Sciences. 2010; 5:161-164.
88. Steel RGD, Torrie JH. Principles and procedures of statistics. Mc. Graw Hill Pub. Co. Inc. New York, 1997.
89. Syed AA, Kadam JA, Mane VP, Patil SS, Baig MMV. Biological efficiency and nutritional contents of *Pleurotus florida* cultivated on different agro-wastes. Natural Science, 2009; 7:44-48.

90. Tan KK. Cotton waste is a good substrate for the cultivation of *P. ostreatus* the oyster mushroom. *Mush. Sci*, 1981; 11:705-10.
91. Tentratian S, Fields ML. Enrichment of ground corn cobs with cellulolytic microorganisms. *Biol. Wastes*, 1990; 34:123-131.
92. Wang D, Sakoda A, Suzuki M. Biological efficiency and nutritional value of *Pleurotus ostreatus* cultivated on spent beet grain. *Bioresources technology*, 2001; 78:293300.
93. Zadrazil F, Brunnert H. Investigation of physical parameters important for solid-state fermentation of straw by white-rot fungi. *Eur. J. Microbiol. Biotechnol.* 1981; 11:183-188.
94. Zadrazil F. Cultivation, yield and keeping quality of *P. florida* Fovo. *Sc. Champignon*, 1973; 13:17-19.
95. Zadrazil F, Change ST, Hayes WA. Cultivation of *Pleurotus* sp. The biology and cultivation of edible mushroom. Academic Press. New York, 1978, 521-555.
96. Zhang RH, Li XJ, Fadel J. Oyster mushroom cultivation with paddy and wheat straw, *Bioresource Technology*, 2002; 82:277-284.