

Vol 5 Issue 12 September 2016

ISSN No : 2249-894X

---

*Monthly Multidisciplinary  
Research Journal*

*Review Of  
Research Journal*

Chief Editors

---

**Ashok Yakkaldevi**  
A R Burla College, India

**Ecaterina Patrascu**  
Spiru Haret University, Bucharest

**Kamani Perera**  
Regional Centre For Strategic Studies,  
Sri Lanka

Review Of Research Journal is a multidisciplinary research journal, published monthly in English, Hindi & Marathi Language. All research papers submitted to the journal will be double - blind peer reviewed referred by members of the editorial Board readers will include investigator in universities, research institutes government and industry with research interest in the general subjects.

### Regional Editor

Dr. T. Manichander

### Advisory Board

Kamani Perera Regional Centre For Strategic Studies, Sri Lanka	Delia Serbescu Spiru Haret University, Bucharest, Romania	Mabel Miao Center for China and Globalization, China
Ecaterina Patrascu Spiru Haret University, Bucharest	Xiaohua Yang University of San Francisco, San Francisco	Ruth Wolf University Walla, Israel
Fabricio Moraes de Almeida Federal University of Rondonia, Brazil	Karina Xavier Massachusetts Institute of Technology (MIT), USA	Jie Hao University of Sydney, Australia
Anna Maria Constantinovici AL. I. Cuza University, Romania	May Hongmei Gao Kennesaw State University, USA	Pei-Shan Kao Andrea University of Essex, United Kingdom
Romona Mihaila Spiru Haret University, Romania	Marc Fetscherin Rollins College, USA	Loredana Bosca Spiru Haret University, Romania
	Liu Chen Beijing Foreign Studies University, China	Ilie Pinte Spiru Haret University, Romania
Mahdi Moharrampour Islamic Azad University buinzahra Branch, Qazvin, Iran	Nimita Khanna Director, Isara Institute of Management, New Delhi	Govind P. Shinde Bharati Vidyapeeth School of Distance Education Center, Navi Mumbai
Titus Pop PhD, Partium Christian University, Oradea, Romania	Salve R. N. Department of Sociology, Shivaji University, Kolhapur	Sonal Singh Vikram University, Ujjain
J. K. VIJAYAKUMAR King Abdullah University of Science & Technology, Saudi Arabia.	P. Malyadri Government Degree College, Tandur, A.P.	Jayashree Patil-Dake MBA Department of Badruka College Commerce and Arts Post Graduate Centre (BCCAPGC), Kachiguda, Hyderabad
George - Calin SERITAN Postdoctoral Researcher Faculty of Philosophy and Socio-Political Sciences Al. I. Cuza University, Iasi	S. D. Sindkhedkar PSGVP Mandal's Arts, Science and Commerce College, Shahada [ M.S. ]	Maj. Dr. S. Bakhtiar Choudhary Director, Hyderabad AP India.
REZA KAFIPOUR Shiraz University of Medical Sciences Shiraz, Iran	Anurag Misra DBS College, Kanpur	AR. SARAVANAKUMARALAGAPPA UNIVERSITY, KARAIKUDI, TN
Rajendra Shendge Director, B.C.U.D. Solapur University, Solapur	C. D. Balaji Panimalar Engineering College, Chennai	V.MAHALAKSHMI Dean, Panimalar Engineering College
	Bhavana vivek patole PhD, Elphinstone college mumbai-32	S.KANNAN Ph.D , Annamalai University
	Awadhesh Kumar Shirotriya Secretary, Play India Play (Trust), Meerut (U.P.)	Kanwar Dinesh Singh Dept.English, Government Postgraduate College , solan

More.....



## BIOCHEMICAL CHANGES IN CARTHAMUS TINCTORIUS L. CULTIVARS UNDER THE PATHOGENESIS OF ALTERNARIA CARTHAMI

Vijay B. Mahadik and N.S. Mali

Department of Botany, Shankarrao Mohite Mahavidyalaya, Akulj. MS (INDIA).

### ABSTRACT

In the present investigation, changes in biochemicals like carbohydrates, chlorophylls, soluble protein and total free amino acids of healthy and infected leaves were analyzed in four cultivars of *Carthamus tinctorius* L. under pathogenesis of *Alternaria carthami*. The biochemicals were decreased in infected leaves as compared to healthy leaves. Carbohydrates and protein content was significantly declined in Nari-6 whereas chlorophylls and total free amino acids were greatly reduced in the infected leaves of Nari-38 and Nari-NH-1 respectively over healthy leaves as compared to other cultivars. The present investigation shows a decrease in carbohydrates, chlorophylls, protein and total free amino acids and also decrease in resistance in all cultivars under the pathogenesis of *Alternaria carthami*. The decrease in disease resistance was confirmed on the basis of morphological observations of the cultivars where they showed increased susceptibility to the pathogen in comparison to healthy cultivars.

**KEYWORDS:** *Carthamus*, *Alternaria*, carbohydrates, protein,

*chlorophylls*, amino acids.

### INTRODUCTION

Safflower (*Carthamus tinctorius*, L.) is one of the important oil seed crops and it is susceptible to number of fungal diseases i.e. *Alternaria* leaf spot (*Alternaria carthami*), *Cercospora* leaf spot (*Cercospora carthami*), *Fusarium* wilt (*Fusarium oxysporum* f.sp. *carthami*), *Ramularia* leaf spot (*Ramularia* spp.), *Rust* (*Puccinia carthami*) etc. Leaf blight caused by *Alternaria carthami* Chowdhury is one of the major diseases of safflower (Deokar et al. 1991). Foliar diseases are most destructive in India which reduces the yield about 90 percent.

The earlier studies carried by Chung (2012) showed that infection caused by *Alternaria* species produces toxin which leads to lipid peroxidation, generation of hydrogen peroxide and followed by cell death. During the course of infection, the fungal toxin produced by the fungus drastically alters the plant photosynthetic network along with physiological and biochemical status (Prakash, 2004; Mohsan et al., 2011; Lin et al., 2011). The role of sugars in plant pathogen interaction has been studied by the earlier workers and it was observed that keto-hexose variation during infection is directly related to the degree of pathogen infection and plant resistance. The studies carried out by Srivastava & Pandey (2012) showed that the total carbohydrate decreased due to fungal infection. The fungal infection usually leads to the chlorosis and necrosis of the leaves which decreases the photosynthetic products (Ciuffetti et al., 2010). Similar study was carried out by Chen et al., (2005) showing the destructive effect of the pathogen on the host



metabolites and on chloroplasts. There is breakdown of the proteins to amino-acids in the host cells during infection, which leads to decrease in protein content was shown by Odebode and Sanusi (1996). There is not enough evidence available on the comparative studies among the biochemical changes of healthy and infected safflower leaves. Hence it was felt necessary to investigation the biochemical changes occurring in *Carthamus tinctorius* L. cultivars under the pathogenesis of *Alternaria carthami* Chowdhury.

## MATERIAL AND METHODS

The seeds of four cultivars of *Carthamus tinctorios* L. like- Nari-38 (spiny variety), Nari-6 (non-spiny variety), Nari-H-23 (spiny hybrid) and Nari-NH-1 (non-spiny hybrid) were collected from Nimkar Agricultural Research Institute, Phaltan, Dist. Satara (MS) and shown in RBD manner in 4 replications during Rabi season of the year 2014 and 2015. The plant protection measures were applied to each replication except one which was treated as controlled (untreated) one (No plant protection measures applied). Healthy and leaves infected with *Alternaria carthami* (leaf blight) were collected respectively from the replication with plant protection measures applied and the controlled replication of each cultivar. Carbohydrates were determined by the method of Nelson (1944), soluble protein by the method of Lowry et al., (1951), chlorophylls and total free amino acids were analyzed by using the method suggested by Sadasivam and Manickam (1996).

## RESULT AND DISCUSSION

The observation Table 1. shows that, reducing sugar and non-reducing, total sugars, starch and total carbohydrates were drastically declined in infected leaves of all cultivars as compared healthy leaves. Nari-H-23 shows least decrease of reducing sugar (34.92%) followed by Nari-38 (51.71%), Nari-NH-1(55.39%) and Nari-6 (71.53%). The cultivar Nari-6 exhibited highest decreases of reducing sugar. The non reducing sugar of infected leaves was declined in all cultivars except Nari-38 which reveals increase in Nonreducing sugar by 20.37% over healthy leaves. Total sugar was significantly declined in Nari-6 (71.3%) followed by Nari-H 23 (38.86%) and Nari-NH-1(31.64%) whereas it was found least decreases in Nari-38 (29.32%). Nari-H-23 showed least decrease in starch (13.43%) followed by Nari-6 (47.73%), Nari-38 (50.07%) and Nari-NH-1(71.01%). The least decrease of total carbohydrate was found in Nari-H-23 (24.07%) followed by Nari-38 (42.1%), Nari-NH-1(53.23%) and Nari-6 (58.68%). Similar declining pattern of carbohydrates were recorded in greengram by Kulkarni et al., (2009) and Sindhan et al. (1999) infected with *Colletotrichum truncatum* (Schw.) Andrus and Moore and *Cercospora* leaf spot respectively. The reduction in carbohydrate contents after infection was due to rapid hydrolysis of sugars during pathogenesis through enzymes secreted by pathogens and subsequent utilization by pathogens for their development (Jaypal and Mahadevan, 1968).

The amount of chlorophyll 'a', 'b' and total chlorophyll of infected leaves was significantly declined in all cultivars over healthy leaves (Table 2). The least decrease in chlorophyll 'a', 'b' and total chlorophyll was found in Nari-6 about 56.01%, 49.76% and 53.44% respectively. Similarly the least decrease in chlorophyll 'a', 'b' and total chlorophyll was found in Nari-H-23 was about 58.36%, 60.2% and 59.7 % respectively. The highest decrease of chlorophyll 'a' was observed in Nari-NH-1 (74.16%) whereas chlorophyll 'b' and total chlorophyll about 96.88% and 83.08% respectively in Nari-38.

Similar findings were observed by Mesta (2006), while working on *Alternaria* blight of sunflower and Kulkarni et al., (2009) in greengram infected with *Colletotrichum truncatum* (Schw.) Andrus and Moore. Alqarawi et al., (2013) showed the significant alterations of chlorophyll 'a' and chlorophyll 'b' in mangrove infected with *Alternaria alternata* (fr.) keissler compared to control plants. Gabara et al., (2012) suggested that decrease of chlorophyll in infected plants may be due to inhibition of photophosphorylation by fungal toxins.

Soluble proteins as well as total free amino acids were also decreased in infected leaves in all cultivars as compared to healthy leaves (Table 3). The least decrease of soluble protein was found in Nari-NH-1 (11.26%) while Nari-6 shows least decrease of total free amino acids (7.01%). The content of soluble protein and total free amino acids were significantly reduced in Nari-6 (31.75%) and Nari-NH-1(65.78%) respectively

as compared to other cultivars. Similar findings were reported in sunflower seed infected with *Sclerotinia* head rot by Kumar et al., (1998) and Thomas and Mathew (2014) in *Lawsonia inermis* L. infected with *Asterina lawsoniae* Henn. & n. This indicates the inhibition of synthesis of amino acids and protein is mainly due to infection. There seems no degradation of protein to free amino acids. The enzymes involved in amino acids synthesis are activated as a result of infection, which lead to decrease in amino acids content in the infected leaves (Rudolph, 1963). The decreased content of soluble protein in the infected leaves is co-ordination with the increase in protease enzymes in the infected leaves was observed by Thomas and Mathew (2014) in *Lawsonia inermis* L. and Pareek and Varma (2015) in cluster bean.

**Table 1. Effect of *Alternaria carthami* infection on carbohydrates in safflower cultivars.**

Cultivars	Parameters	Carbohydrates mg g <sup>-1</sup> fresh weight				
		Reducing sugar	Non-reducing sugar	Total sugar	Starch	Total carbohydrates
Nari-38	Healthy leaves	14.61±0.35	6.58±0.56	21.2±0.22	34.01±0.21	55.21±0.41
	Infected leaves	7.05±0.21 (51.71)*	7.92±1.08 (20.37)*	14.98±0.87 (29.32)*	16.98±1.62 (50.07)*	31.96±0.75 (42.1)*
Nari-6	Healthy leaves	10.15±0.55	8.51±0.47	18.67±0.87	21.50±0.29	40.18±1.16
	Infected leaves	2.89±0.24 (71.53)*	2.46±0.48 (71.03)*	5.35±0.69 (71.3)*	11.24±0.08 (47.73)*	16.6±0.69 (58.68)*
Nari-H-23	Healthy leaves	24.65±5.80	6.04±3.25	30.69±2.66	42.68±0.40	73.37±2.37
	Infected leaves	16.04±0.08 (34.92)*	2.72±0.33 (54.95)*	18.76±0.39 (38.86)*	36.95±1.52 (13.43)*	55.71±1.17 (24.07)*
Nari-NH-1	Healthy leaves	10.2±0.84	11.52±1.01	21.72±0.22	26.39±3.45	48.11±3.61
	Infected leaves	4.55±1.02 (55.39)*	10.3±1.28 (10.62)*	14.85±0.40 (31.64)*	7.65±0.46 (71.01)*	22.50±0.44 (53.23)*

(Values are mean ± SD of triplicates. \* Per cent increase or decrease over Healthy leaves)

**Table 2. Effect of *Alternaria carthami* infection on chlorophylls in safflower cultivars.**

Cultivars	Parameters	chlorophylls mg g <sup>-1</sup> fresh weight		
		Chlorophyll 'a'	chlorophyll 'b'	Total chlorophyll
Nari-38	Healthy leaves	9.944±0.184	9.216±0.180	19.161±0.241
	Infected leaves	2.954±0.095 (70.28)*	0.287±0.089 (96.88)*	3.242±0.180 (83.08)*
Nari-6	Healthy leaves	9.215±0.105	6.426±0.277	15.641±0.351
	Infected leaves	4.053±0.157 (56.01)*	3.228±0.235 (49.76)*	7.281±0.295 (53.44)*
Nari-H-23	Healthy leaves	9.914±0.072	6.233±0.247	16.148±0.215
	Infected leaves	4.127±0.167 (58.36)*	2.48±0.210 (60.2)*	6.608±0.289 (59.07)*
Nari-NH-1	Healthy leaves	14.037±0.074	9.361±0.169	23.399±0.147
	Infected leaves	3.626±0.104 (74.16)*	2.419±0.196 (74.15)*	6.046±0.293 (74.15)*

(Values are mean ± SD of triplicates. \* Per cent increase or decrease over Healthy leaves)



**Table 3. Effect of *Alternaria carthami* infection on soluble protein and total free amino acids in safflower cultivars.**

Cultivars	Parameters	Soluble protein mg g <sup>-1</sup> fresh weight	Total free amino acids mg g <sup>-1</sup> fresh weight
Nari-38	Healthy leaves	35.89±0.57	13.70±0.82
	Infected leaves	28.67±0.61 (20.12%)*	7.58±0.41 (44.68%)*
Nari-6	Healthy leaves	33.30±0.32	16.62±0.71
	Infected leaves	22.73±0.59 (31.75%)*	15.45±4.06 (7.01%)*
Nari-H-23	Healthy leaves	35.54±0.13	22.45±0.82
	Infected leaves	30.62±0.65 (13.83%)*	9.04±1.79 (59.74%)*
Nari-NH-1	Healthy leaves	26.76±0.65	22.16±2.29
	Infected leaves	23.75±0.63 (11.26%)*	7.58±0.82 (65.78%)*

(Values are mean ± SD of triplicates. \* Per cent increase or decrease over Healthy leaves)

Based on the present findings, it is concluded that the Nari-6 and Nari-H-23 exhibit least decrease of biochemicals in infected leaves making them more resistant whereas Nari-38 and Nari-NH-1 show highest decrease of biochemicals in infected leaves which makes them more susceptible to *Alternaria carthami* infection.

#### ACKNOWLEDGMENTS

The authors acknowledge laboratory facilities provided by Principal, Shankarrao Mohite Mahavidyalaya, Akhuj during experimental work.

#### REFERENCES

- Alqarawi, A. A., Hashem, A and Abd-allah, E. F. (2013). Physiological alterations in *Avicennia marina* (forski) vierh associated with leaf spot disease caused by *Alternaria alternata* (fr.) keissler. Pak. J. Bot., 45(5): 1767-1771.
- Chen, S., Dai, X., Qiang, S and Tang, Y. (2005). Effect of a nonhost-selective toxin from *Alternaria alternata* on chloroplast-electron transfer activity in *Eupatorium adenophorum*. Plant Pathol., 54: 671-677.
- Chung, K. R. (2012). Stress Response and Pathogenicity of the Necrotrophic Fungal Pathogen *Alternaria alternata*. Scientifica, Article ID 635431, 17 pages. <http://dx.doi.org/10.6064/2012/635431>.
- Ciuffetti, L. M., Manning, V. A., Pandelova, I., Betts, M. F and Martinez, J. P. (2010). Host-selective toxins, Ptr ToxA and Ptr ToxB, as necrotrophic effectors in the *Pyrenophora tritici-repentis*-wheat interaction. New Phytol., 187: 911-919.
- Deokar, C. D., Veer, D. M., Patil, R. C and Ranga RAO, V. (1991). Survey of safflower diseases in Maharashtra State. Sesame and Safflower News letter, 6: 79-80.
- Gabara, B., Kzhiak, E., Sklodowska, M., Surowka, E and Miszalski, Z. (2012). Ultra structural and metabolic modifications at the plant pathogen interface in *Mesembryanthemum crystallinum* leaves infected by *Botrytis cinerea*. Environ Exp Bot., 77:33.
- Jayapal, R and Mahadevan, A. (1968) Biochemical changes in banana leaves in response of leaf spot pathogens. Indian Phytopathol., 21: 43-48.
- Kulkarni, S., Benagi, V. I., Patil, . P. V., Hegde, Y., Konda, C. R and Deshpande, V. K. (2009). Sources of

- resistance to anthracnose in greengram and biochemical parameters for resistance. *Karnataka J. Agric. Sci.*, 22 (5): 1123-1125.
9. Kumar, B., Chahal, S. S and Ahuja, K. L. (1998). Effect of *Sclerotinia* head rot on some bioconstituents of sunflower seed. *Indian Phytopath.* 51 (4): 359-360
10. Lin, C. H., Yang, S. L and Chung, K. R. (2011). Cellular responses required for oxidative stress tolerance, colonization, and lesion formation by the necrotrophic fungus *Alternaria alternata* in citrus. *Curr. Microbiol.*, 62(3): 807-815.
11. Lowry, O. H., Rosenbrough, N. J., Farr, A. L and Randall, R. J. (1951). Protein measurement with folin phenol reagent. *J. Biol. Chem.*, 193:262-263.
12. Mesta, R. K. (2006). Epidemiology and management of *Alternaria* blight of sunflower caused by *Alternaria helianthi* (Hansf.) Tubaki and Nishihara. Ph. D. Thesis, Univ. Agric. Sci., Dharwad (India).
13. Mohsan, M. M., Intizar-ul-Hassan, Liaquat, A. (2011). Chemotherapeutic management of *Alternaria* black spot (*Alternaria alternata*) in mango fruits. *J. Agric. Res.*, 49: 499-505.
14. Nelson, N. (1944). Photometric adaptation of the somogyi method for the determination of glucose. *J. Biol. Chem.*, 153:379-380.
15. Odebode, A. C and Sanusi, J. (1996): Influence of fungi associated with banana on nutritional content during storage. *Z. Lebensum unters Forsch.* 202: 471-473.
16. Pareek, V and Varma, R. (2015). Estimation of protein and its related enzyme in cluster bean plant parts infected with *Fusarium solani* caused wilt disease. *International Journal of Multidisciplinary Research and Development.* Vol. 2 (11): 241-244
17. Prakash, O. M. (2004). Disease and disorders of Mango and their Management. In: *Disease of Fruits and Vegetables.* Vol. 1. S. A. M. H. Naqvi (eds.). Academic Publishers Netherlands. pp. 511-619.
18. Rudolph, K. (1963). Weitere biochemische Untersuchungen zum Wirt-parasitenverhältnis am Beispiel von *Puccinia graminis tritici*. *Phytopathol.* 46: 276-290.
19. Sadashivam, S and Manikam, A. (1996). "Biochemical methods", Second Edn. Wiley Eastern, Delhi. pp-250.
20. Sindhan, G. S., Indra Hooda and Parashar, R. D. (1999). Sources of resistance of *Cercospora* leaf spot in mungbean and biochemical parameters for resistance. *J. Mycol. Pl. Path.*, 29: 130-132.
21. Srivastava, A. K and Pandey, G. K. (2012). Starch and glucose content and changes therein in the seeds of *Schleichera oleosa* (kusum) due to bioteriation by pathogenic fungi during storage. *Int. J. Recent Trends Sci. Tech.*, 4: 5-8.
22. Thomas, J., and Mathew, K. L. (2014). Biochemical changes in the *Lawsonia inermis* infected with *Asterina lawsoniae* Henn. & Nyn. *Current Research in Environmental & Applied Mycology.*, 4(2): 217-220.



**Vijay B. Mahadik**  
Department of Botany, Shankarrao Mohite  
Mahavidyalaya, Akulj. M S (INDIA).

# Publish Research Article

## International Level Multidisciplinary Research Journal For All Subjects

Dear Sir/Mam,

We invite unpublished Research Paper, Summary of Research Project, Theses, Books and Books Review for publication, you will be pleased to know that our journals are

### Associated and Indexed, India

- ★ Directory Of Research Journal Indexing
- ★ International Scientific Journal Consortium Scientific
- ★ OPEN J-GATE

### Associated and Indexed, USA

- DOAJ
- EBSCO
- Crossref DOI
- Index Copernicus
- Publication Index
- Academic Journal Database
- Contemporary Research Index
- Academic Paper Database
- Digital Journals Database
- Current Index to Scholarly Journals
- Elite Scientific Journal Archive
- Directory Of Academic Resources
- Scholar Journal Index
- Recent Science Index
- Scientific Resources Database

Review Of Research Journal  
258/34 Raviwar Peth Solapur-  
413005, Maharashtra  
Contact-9595359435

E-Mail-ayisrj@yahoo.in/ayisrj2011@gmail.com