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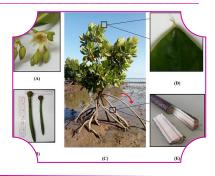
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ANTIBACTERIAL ACTIVITY OF MANGROVE PLANT RHIZOPHORA MUCRONATA LAMK. FROM THE COAST OF RAIGAD DISTRICT, MAHARASHTRA

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ABSTRACT

The objective of present study is to investigate the antibacterial activities of n-hexane, ethyl acetate and methanol extracts of the leaves of Rhizophora mucronata Lamk. against six human pathogenic microbes. The antibacterial activity was evaluated using disc diffusion and microdilution methods. The antibacterial activities of the crude extracts were increased with creasing in the concentration. It is clear that n-hexane extract was the most effective extract. Additionally, gram positive Bacillus cereus (B. cereus) appear to be the most sensitive strain while Pseudomonas aeruginosa (P. aeruginosa). The inhibition of bacterial growth at concentration as low as 0.04 mg/mL indicated the potent antibacterial activity of R. mucronata extracts. This concludes that the obtained results are considered sufficient for further study to isolate the compounds responsible for the antibacterial activity and suggesting the possibility of finding potent antibacterial agents from R. mucronata extracts.

KEYWORDS: Mangoves, Rhizophora mucronata Lamk., antibacterial activity, Raigad.

1. INTRODUCTION

Infectious diseases represent a serious public health problem and they remain the leading cause of death throughout the world^[1-3]. Currently, the problems of microbial drug resistance, an increase of opportunistic infections and the toxicity effect of continued use of several antimicrobial drugs^[4] have necessitated a search for new antimicrobial drugs from other sources including natural sources like plants which are the good sources of novel antimicrobial chemotherapeutic agents. Furthermore, plants have been a major source for drug development^[5-7]. Plant extracts and products are used in the treatment of infectious disease^[8-10]. *Rhizophora mucronata* Lamk. (*R. mucronata*) -locally known as *Kandal*- is large mangrove tree (up to 10 - 12 m tall) belongs to the Rhizophoraceae family. It is distributed throughout the Indian coast ^[11]. However, *R. mucronata* has been known traditionally as an important remedy for sprue^[11]. Up to date, there are no study has been conducted on the evaluation of the antibacterial activity of this plant. Therefore, this study aims to investigate the antimicrobial activities of *R. mucronata* extracts against four human pathogenic bacteria including two Gram-positive (*Staphylococcusa ureus* (*S. aureus*) ATCC25923, *Bacillus cereus* (*B. cereus*) ATCC11778) and two Gram-negative (*Pseudomonas aeruginosa* (*P. aeruginosa*) ATCC27853, *Escherichia coli* (*E. coli*) ATCC35218). The efficacy of n-hexane, ethyl acetate and methanol extracts from the leaves of *R. mucronata* were also investigated and described.

2. MATERIALS AND METHODS

2.1. Plant Collection

R. mucronata was collected from Maharashtra coast at Raigad district in August 2017. The Voucher of the specimen was deposited in the Department of Botany, Doshi Vakil Arts College and G.C.U.B. Science & Commerce College, Goregaon-Raigad. The taxonomic identification of this plant was done by using Flora of Maharashtra.

2.2. Plant preparation and extraction

The fresh plant was washed under running tap water and dried in a warm room for 3 to 5 d. The samples were grinded into fine powder and extracted by Soxhlet with n-hexane, ethyl acetate and methanol successively to get n-hexane, ethyl acetate and methanol extracts. Then, all the crude was kept at -20 °C until further use.

2.3. Samples preparation

A sample of 100 mg from each extract was dissolved in1 mL DMSO. The extract was then sterilized by filtration through sterile syringe filter (0.2 μ m pore). Finally the filtered extract was stored as aliquots until it was used.

2.4. Microbial strains

Four reference strains of human pathogenic bacteria were used in this study including two Grampositive (*S. aureus* ATCC25923, *B. cereus* ATCC11778) and two Gram-negative (*P. aeruginosa* ATCC27853, *E. coli* ATCC35218).

2.5. Antimicrobial assay

2.5.1. Disc diffusion method

The agar disc diffusion method was employed for the determination of antibacterial activities of the extracts according to Qaralleh *et al.*^[12] with some modification. Briefly, inoculum containing 107 CFU/mL was spread on Mueller-Hinton agar plates for four bacteria. Using sterile forceps, the sterile filter papers (6 mm diameter) containing the crude extracts (1 or 1.5 mg), standard antibiotics (30 μ g of chloramphenicol) or negative control (DMSO) were laid down on the surface of inoculated agar plate. The plates were incubated at 37 °C for 24 hours. Each sample was tested in duplicate and the zone of inhibition was measured as millimeter diameter.

2.5.2. Microdilution method

Minimum inhibitory concentration (MIC) was measured by determining the smallest amount of extract or standard antibiotic needed to inhibit the visible growth of a test bacterium. This was done using 96-well plates. The assay plates were filled with Mueller- Hinton broth medium (MHB) containing different concentrations of extracts, tetracycline or solvent control and the test bacterium (107 CFU/mL). Each sample was tested in triplicate and the observation was recorded by naked eyes after 24 h incubation periods at 37 °C. Minimal bactericidal concentration (MBC) was determined by transferring and spreading the treated culture broth of the wells containing the concentrations equal to and higher than the MIC on agar plates. The lowest concentration of the extract or the standard antibiotic required to completely destroy test bacteria after incubation at 37 °C for 24 h was reported as MBC.

3. RESULTS

The antibacterial activity of *R. mucronata* extracts are shown in Table 1. Generally, the results showed that the antibacterial activities of the crude extracts were increased with increasing the concentration. Although the antibacterial activity of the extracts tested is variable, two Gram-positive bacteria (*S. aureus* and *B. cereus*) and only gram negative (*E. coli*) were inhibited by the extracts.

Quantitative analyses on the antibacterial properties were obtained through the determination of bacteriostatic and bactericidal concentrations of R. mucronata extracts. Table2 shows the MIC and MBC of the extracts that produce inhibition zone more than 12 mm. The results of inhibition zone were reflected in lower MIC values. The MIC and MBC values for bacterial strains, which sensitive to the extracts, were in the range of 0.04-1.11 mg/mL and 0.04-10 mg/mL, respectively. Furthermore, in most cases, the MBC values were higher than the MIC values, except for n-hexane extract against *B. cereus* (MIC = MBC). According to the disc diffusion results, MIC and MBC values, it is clearthat n-hexane extract was the most effective extract (Table 1 and 2). Additionally, Gram positive *B. cereus* appears to be the most sensitive strain with inhibition zone of 19 mm (1.5 mg/disc) and the MIC value is 0.04 mg/mL). The inhibition of microbial growth at concentration as low as 0.04 mg/mL indicated the potent antibacterial activity of R. mucronata extracts.

Table 1: Antimicrobial activity of R. mucronata extracts

Microorganisms	Zone of Inhibition n-hexane		Ethyl acetate		Methanol		Positive	Negative
	1 mg	1.5 mg	1 mg _	1.5 mg	1 mg	1.5 mg	control	control
S. aureus	14.5	14.5	11	16	13	17	21	00
B. cereus	16.00	19	09	09	9	9.5	27	00
E. coli	11.5	13.5	13.5	16.5	9.5	11	30.5	00
P. aeuroginosa	00	00	00	00	00	00	30.5	00

Positive control: tetracycline (100 µg); Negative control: DMSO.

	Table 2: MIC					
Extracts	S. aureus		B. cereus		E. coli	
	MIC	MBC	MIC	MBC	MIC	MBC
n-hexane	1.11	>10	0.04	0.04	0.04	0.37
Ethyl acetate	0.37	>10	-	-	0.37	>10.00
Methanol	1.11	10	-	-	-	-
Tetracycline	30.0	-	2.00	-	30.0	-

All data were expressed as (mg/mL) except for tetracycline (µg/mL); -: not determined.

4. DISCUSSION

Traditionally, plants were known as the main sources for drugs. Interest in this area continues and many new potent drugs have been isolated. Tropical and sub-tropical are as of the world are rich with many plant species which have effective properties, such as antimicrobial, antiviral and anti-fungal. Many medicine plant extracts have been known to possess antibacterial effects. Mangroves possess novel biologically active compounds. The extracts from different mangrove plants and mangrove associates have been reported to possess inhibition action against human and plant pathogens^[13-22]. In this report, three different polarity extracts have been tested for antimicrobial activity. With respect to the inhibition panel and the MIC and MBC concentrations, n-hexane extract of *R. mucronata* was the most effective extract. The methanol and dichloromethane extracts of *R. mucronata* also demonstrated antibacterial effect, although they were lower than the antibacterial effects of the n-hexane extract. The presence of more than one active compound. Chemical analysis of the species belongs to the genus *Rhizophora* have shown the presence of various bioactive ingredients including alkaloids, steroids, triterpenoids and flavonoids^[14,23]. On the other hand, it is interesting to note that the plant extracts showed bacteriostatic and bactericidal actions against *S. aureus, B. cereus* and *E. coli*. This suggests that they may possess remarkable therapeutic action in

the treatment of infectious disease caused by these species. The obtained results suggest the possibility of finding potent antibacterial agents from *R. mucronata* extracts and considered sufficient to isolate the compounds responsiblefor the activity.

Conflict of interest statement: I declare that I have no conflict of interest.

REFERENCES:

- Fonkwo PN. Pricing infectious disease: the economic and health implications of infectious diseases. EMBO reports 2008; 9: 13-17.
- Creager ANH. Adaptation or selection? Old issues and new stakes in the postwar debates over bacterial drug resistance. Stud Hist Phil Biol Biomed Sci2007; 38: 159-190.
- Jacoby TS, Kuchenbecker RS, Dos Santos RP, Magedanz L, Guzatto P, Moreira LB. Impact of hospital-wide infection rate, invasive procedures use and antimicrobial consumption on bacterial resistance inside an intensive care unit. J Hos Infect2010; 75: 23-27.
- Sasidharan S, Prema B, Yoga LL. Antimicrobial drug resistance of Staphylococcus aureus in dairy products. Asian Pac J Trop Biomed2011; 1(2): 130-132.
- Soh PN, Benoit-Vical FO. Are West African plants a source of future antimalarial drugs? J Ethnopharmacol2007; 114: 130-140.
- Saklani A, Kutty SK. Plant-derived compounds in clinical trials. Drug Discovery Today 2008; 13(3/4): 161-171.

Pan L, Chai H, Kinghorn AD. The continuing search for anti-tumor agents from higher plants. PhytochemLett2010; 3: 1-8.

- Qaralleh HN, Abboud MM, Khleifat KM, Tarawneh KA, Althunibat OY. Antibacterial activity in vitro of Thymus Capitatus from Jordan. Pak J Pharm Sci2009; 22(3): 247-251.
- Kuete V, Ngameni B, Simo FCC, Tankeu RK, Ngadjui BT, Meyer JJM, et al. Antimicrobial activity of the crude extracts and compounds from Ficus chlamydocarpa and Ficus cordata (Moraceae). J Ethnopharmacol2008; 120: 17-24.
- Lee OK, Lee BY. Antioxidant and antimicrobial activities of individual and combined phenolics in Oleae uropaea leaf extract.BioresourTechnol2010; 101: 3751-3754.
- Manivannan K, Karthikai DG, Anantharaman P, Balasubramanian T. Antimicrobial potential of selected brown seaweeds from Vedalai coastal waters, Gulf of Mannar. Asian Pac J Trop Biomed2011; 1(2): 114-120.
- Qaralleh H, Idid S, Saad S, Susanti D, Taher M, Khleifat K. Antifungal and antibacterial activities of four Malaysian spongespecies (Petrosiidae). J Med Mycol2010; 20(4): 315-320.
- Chandrasekaran M, Kannathasan K, Venkatesalu V, Prabhakar K. Antibacterial activity of some salt marsh halophytes and mangrove plants against methicillin resistant Staphylococcus aureus. World JMicrobiolBiotechnol2009; 2: 155-160.
- Abeysinghe P. Antibacterial activity of some medicinal mangroves against antibiotic resistant pathogenic bacteria. Indian J PharmSci2010; 72(2): 167-172.
- Paul RK, Irudayaraj V, Johnson M, Patric RD. Phytochemical and anti-bacterial activity of epidermal glands extract of Christella parasitica (L.) H. Lev. Asian Pac J Trop Biomed 2011; 1(1): 8-11.
- Sivaperumal P, Ramasamy P, Inbaneson S, Ravikumar S. Screening of antibacterial activity of mangrove leaf bioactive compounds against antibiotic resistant clinical isolates. World J Fish Mar Sci2010; 2(5): 348-353.
- Vadlapudi VR, Bobbarala V. In vitro antimicrobial activity of two mangrove plants Aegiceras corniculatum and Hibiscus tiliaceous. Biosci Biotechnol Res Asia 2009; 6(1): 321-324.
- Ravikumar S, Muthuraja M, Sivaperumal P, Gnanadesign M. Antibacterial activity of the mangrove leaves Exoecaria agallocha against selected fish pathogens. Asian J Med Sci2010; 2(5): 211- 213.
- Huang Z, Cai Z, Shao C, She Z, Xia X, Chen Y. et al. Chemistry and weak antimicrobial activities of phomopsins produced by mangrove endophytic fungus Phomopsissp. ZSU-H76.Phytochem2008; 69: 1604-1608.

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- Hu HQ, Li ZS, He H. Characterization of an antimicrobial material from anewly isolated Bacillus amylo liquefaciens from mangrove for biocontrol of Capsicum bacterial wilt. Biol Control 2010; 54: 359-365.
- Marrero E, Sánchez J, Armas ED, Escobar A, Melchor G, Abad MJ, et al. COX-2 and sPLA2 inhibitory activity of aqueous extract and polyphenols of Rhizophora mangle (red mangrove). Fitoterapia2006; 77(4): 313-315.
- Khajure PV, Rathod JL. Antimicrobial activity of extracts o facanthus ilicifolius extracted from the mangroves of karwar coast karnataka. Rec Res SciTechnol2010; 2(6): 98-99.
- Mandal MD, Mandal S. Honey: its medicinal property and antibacterial activity. Asian Pac J Trop Biomed 2011; 1(2):154-160.
- Habbal O, Hasson SS, El-Hag AH, Al-Mahrooqi Z, Al-Hashmi N, Al-Bimani Z, et al. Antibacterial activity of Lawsonia inermis Linn (Henna) against Pseudomonas aeruginosa. Asian Pac J Trop Biomed 2011; 1(3): 173-176.
- Tirupathi RG, Suresh BK, Kumar JU, Sujana P, Rao AV, Sreedhar AS. Anti-microbial principles of selected remedial plants from Southern India. Asian Pac J Trop Biomed 2011; 1(4): 298-305.
- Taye B, Giday M, Animut A, Seid J. Antibacterial activities of selected medicinal plants in traditional treatment of human wounds in Ethiopia. Asian Pac J Trop Biomed 2011; 1(5):370-375.
- Elumalai EK, Ramachandran M, Thirumalai T, Vinoth kumar P. Antibacterial activity of various leaf extracts of Merremiae marginata. Asian Pac J Trop Biomed 2011; 1(5): 406-408.
- Kader G, Nikkon F, Rashid MA, Yeasmin T. Antimicrobial activities of the rhizome extract of Zingiber zerumbet Linn. Asian Pac J Trop Biomed 2011; 1(5): 409-412.
- Prasad TNVKV, Elumalai EK. Biofabrication of Ag nano-particles using Moringao leifera leaf extract and their antimicrobial activity. Asian Pac J Trop Biomed 2011; 1(6): 439-442.
- Khan AV, Ahmed AU, Mir MR, Shukla I, Khan AA. Antibacterial efficacy of the seed extracts of Melia azedarach against some hospital isolated human pathogenic bacterial strains. Asian Pac J Trop Biomed 2011; 1(6): 452-455.