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HISTOLOGICAL ALTERATIONS IN GILL TISSUES OF ANABAS TESTIDUNEUS ON EXPOSURE TO FERTILIZER DAP

Avdhesh Kumar Thakur
Research Scholar , LNMU, Darbhanga.

ABSTRACT

*Histological alterations are the variations arising in the tissues of the organisms after exposure to certain chemicals found in the ecosystem. These alterations may be in any part or organs of the organisms and have been studied extensively. Biological systems are open to fertilizer DAP exposure in the environment. Various researchers have found cellular and genetic changes in the tissues of organisms more specifically on fishes. Fishes survive in close interaction with the water through their gills and thus susceptible to fertilizer drained from various sources. In the current study an attempt has been made to assess the impact of DAP on the gill tissues of *Anabas testudineus*. The structural changes in the tissues were noticed. Sub lethal concentration of DAP could disturb growth rate and reproduction causing community disturbances in the tropic levels of food chains. Further, computational genoproteomic studies may shed more light on the general ecophysiology of the fishes.*



KEY WORDS: Histology, fish, gill tissues, Fertilizer, DAP.

1. INTRODUCTION

Histological alterations are the unwanted changes found in the tissues of animals after exposure towards certain fertilizer DAP. Due to their toxicity, accumulation and biomagnifications in water, sediment, and in aquatic food chain [1] along with their association with various diseases [2], these fertilizer DAP leads to significant environment hazards for aquatic bodies. Fishes being an important source of food are of interest because these are rich in vitamins, calcium, phosphorous and iodine [3] and are considered as a good indicator for fertilizer contamination because they occupy different trophic levels; are of different sizes and ages as compared to invertebrates and are also more sensitive to many toxicants [4,5,6]. Gills of fishes are the body parts for gaseous exchange and accomplish osmoregulation, acid-base balance and nitrogenous waste excretion [7,8]. The continuing increase of toxic materials more specifically in water due to run off from industries and agriculture have serious impact on the aquatic animals [9]. Thus, the studies on the accumulation of DAP in various organs of the fish help in determining the extent of pollution and their causative harmful effects [6,10]. In the current study an attempt has been made to assess the impact of DAP on the gill tissues of *Anabas testudineus*.

2. MATERIALS AND METHODS

Study area

The present study was carried out in Darbhanga District of Bihar. Darbhanga district is one of the thirty-eight districts of Bihar state in eastern India, and Darbhanga city is the administrative headquarters of this district and 5th largest city of Bihar as well. Darbhanga district is a part of Darbhanga Division. The district is bounded on the north by Madhubani district, on the south by Samastipur district, on the east by Saharsa district and on the west by Sitamarhi and Muzaffarpur districts.

Sample collection

For the current study, live and healthy *Anabas testudineus* of uniform size were collected from the non-polluted area of the Darbhanga district along with the water samples. The fishes were reared and maintained in the laboratory condition in the dechlorinated tap water and no diet was given to them. (Fig. 1)



Fig 1: *Anabas testudineus*: Sample fish species for present Research work

Exposure to DAP

The fishes were categorized into two groups; one group contained the normal fish that is the control group fishes whereas the other group contained the treated fishes. The treated group was exposed to the fertilizer DAP for 24 hours for the study of histological alterations found in them. Histological preparations After 24 hrs of exposure to DAP, the gill samples of both control and treated group of fishes were excised, rinsed with deionized water and kept for preservation by using 10% neutral buffered formalin as the compound fixative for histological processing. After 24 hours the samples were washed under tap water and the tissues were preserved in 70% alcohol. The tissues were dehydrated by using 90% and 100% alcohols. The tissues were kept in xylene for few minutes for clearing and then transferred to the mixture of xylene and paraffin wax for about 30 minutes. During the hot infiltration the tissues were soaked in molten wax for impregnation at a standard temperature coinciding with the melting point of the embedding paraffin wax medium used. This was achieved by passing of the cleared tissue through changes of paraffin wax molten at coinciding melting temperature of wax in each case. The final processing stage was the embedding of tissues in paraffin wax which was necessary to hold the tissue in position and ensure that tissues were not crumbled during sectioning. Blocks were prepared and kept overnight. Then the tissues were trimmed and sections were made using a microtome. For the successful attachment of tissue section along with wax ribbon the cleaned slides were rubbed with bovine serum albumin which was used as adhesive and then the sections were placed on those slides, kept on hot plate for stretching. After that the slides were dried and kept overnight so that those will be ready for staining. Sections were deparaffinized in xylene for about 20 minutes and treated with different grades of alcohol i.e. 100%, 90%, 70%, 50% and 30% respectively. The slides were dipped in water, stained in haematoxylin, washed under running tap water, dehydrated via graded alcohol to 90%, counterstained in alcoholic eosin, rinsed in 90% alcohol, dehydrated in absolute alcohol, cleared in xylene and mounted in DPX.

3. RESULTS AND DISCUSSION

Examination of thin sections of gill arch of *Anabas testudineus* (control) showed four pairs of typical gill arches bearing two rows of primary gill filaments. Each gill filament bears series of alternate ly arranged semicircular secondary lamellae on both sides. The surface of gill lamella was lined by a thin layer of simple squamous epithelium which rests on basemebnt membra ne covering the pillar cell- blood channel system and which constitutes the main vascular area of the gill. There are several reports on the types of histological changes in fish gills due to contaminate d water, in field and after acute or chronic exposure in la boratory conditions with sub lethal and lethal concentration fertilizer like DAP phosphate etc. [11-20]. In the present work, gills of *Anabas testudineus* exposed to DAP solution exhibited varying degree of damage in sub lethal concentrations. Cell hyperplasia was generally more pronounced towards the proximal end of the filament. After hrs of exposure, hyperplasia of epithelial cells resulted in the fusion of many lamellae. The control group of fishes contain normal gill lamella, gill bar and epithelial cell which were clearly seen whereas in group fishes the gill tissue were greatly affected like necrosis took place, hyperplasia has been noticed which has perhaps damaged the gill lamella (Fig. 2&3).



Fig 2. Gill of *Anabas testudineus* (control); EC : endothelial cell, L: lamella

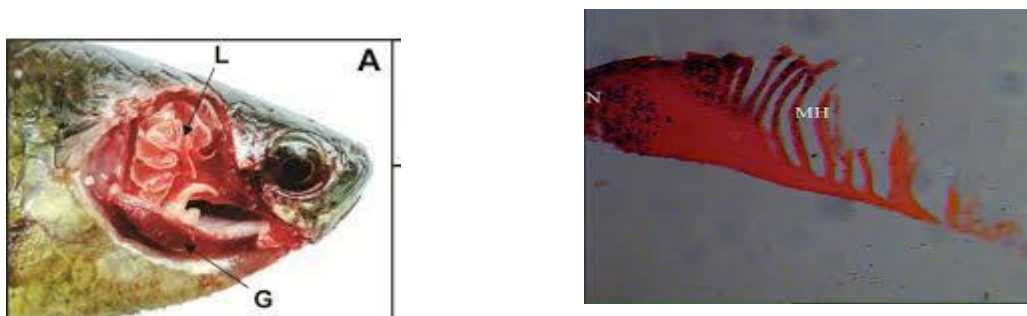


Fig 3: Gill of *Anabas testudineus* (treated); N: Necrosis, HP: hyperplasia. MH: mild

Gills have widespread surface area, blood capillaries for efficient gaseous exchange and provided with mucus cells [21]. The mucous discharge works against toxic substances. Due to DAP intoxication the gill epithelium was compeletely separated from the basement membrane and pillar cells and there was a swelling of the secondary lamellae and dilation of the vessels. The pillar cell nucleus showed necrosis and vacuolation in the secondary gill epithelium. The disorganized fusion in secondary gill epithelium was prominently noticed. Similar histological alterations in the gills were noticed by Velmurugan and coworkers [22] after exposure to organophosphates leadin o epithelial proliferation, congestion of blood vellse and hyperplasia of mucus cells. The physical changes in the gills have been studied leading to necrosis, rupture of the bra nchial epithelium, autolysis, swelling and lamellar fusion. The accumulations of the DAP decrease ventilation which ultimately decreased the O2 uptake. Similar finding was reptre d by Prashanth and others [23] leading to epithelial lifting in the Nile tilapia (*Oreochromis niloticus*) under exposure to glyphosate for 96h. The enlargement of chloride secreting cells and their nuclei supports the above assumption. One of the

important observations in the present study was the fusion of secondary lamella. This could be attributed to counter stress and transformation of electrically charged properties of the epithelial cells which favor adhesion between the cells of two neighboring secondary lamellae. The fusion of secondary lamellae causing a drastic reduction in the respiratory surface area. DAP could have induced fusion of secondary lamella of gills. Hence it could be assumed that copper sulphate intoxication caused severe aerobic stress in *Anabas testudineus* leading to wear and tear in the gill epithelium [24]. The other variations in the gill epithelium were the separation of respiratory epithelium from basement membrane leading to increasing thickness of secondary lamella thereby decreased diffusion capacity and forming a barrier to prevent entering of dissolved DAP. Identical lifting of the respiratory epithelium of secondary lamella of the gills has also been observed in *H. fossilis* subjected to desiccation stress [25].

4. CONCLUSION

DAP causes toxic effects on gill tissues leading to structural deformations. In sub lethal concentration it may be fatal for the organisms affecting the growth rate and reproduction resulting in disturbance to whole community and also trophic levels of food chains, ultimately the ecosystem. Further, computational genoproteomic studies may shed more light on the general exophysiology of the fishes.

REFERENCES

1. Adeogun AO. Impact of industrial effluent on water quality and gill pathology of *Clarias gariepinus* from Alaro stream, Ibadan, Southwest, Nigeria. *European Journal of Scientific Research*. 2012; 76 (1) 83-94
2. Uwem GU, Emile AF, Udo IJ, et al. Bioaccumulation of heavy metal in three fresh water fishes caught from cross river system. *European Journal of Experimental Biology*. 2013; 3(3): 576-582.
3. Zaki MS, Shalaby SI, Ata N, et al. Effect of Aquatic Pollution on Fish (Review). *Life science journal* 2013 ; 10(1): 637-642.
4. Powers DA. Fish as model systems. *Science*. 1989; 246(4928): 352-3588.
5. Wester PW, Vethaak AD, van Muiswinkel WB. Fish as biomarkers in immunotoxicology. *Toxicology* 1994; 83(3): 213-232.
6. Yancheva V, Velcheva I Stoyanova S, et al. Histological Biomarkers in fish as a tool, Ecological risk assessment and monitoring programs: A Review. *Applied Ecology and Environmental Research*. 2015; 14(1) : 47-75
7. Heath AG. *Water pollution and fish physiology*. CRC Press, Florida. 1987.
8. Hadi AA, Alwan SF. Histopathological changes in gills, liver and kidney of fresh water fish, *Tilapia zillii*, exposed to aluminum. *International Journal of Pharmacy & Life Sciences*. 2012; 3(11): 2071-2081.
9. Dutta HM, Munshi JSD, Roy PK, Singh NK, Adhikari S-Ultrastructural in the respiration lamellae of the catfish *Heteropneustes fossilis* after sub lethal exposure of malathion. *Environ Pollut* 1996; 3: 329-341.
10. Shivakumar CK, Thippeswamy B, Tejaswi kumar MV, et al. Bioaccumulation of DAP and its effect on organs of edible fishes located in Bhadra River, Karnataka. *International Journal of Research in Fisheries and Aquaculture*. 4(2): 90-98
11. Doughtie DG, Rao KR. Ultrastructural and histological study of degenerative changes leading to black gills in grass shrimp exposed to a dithiocarbamate biocide. *Journal of Invertebrate Pathology* 1983; 41(1):33 -50.
12. Alazemi BM, Lewis JW, Andrews EB. Gill damage in the freshwater fish *Gnathonemus petersi* (Family: Mormyridae) exposed to selected pollutants : an ultrastructural study. *Environmental Technology* 1996; 17(3): 225-238.
13. Mazon AF Monteiro EAS, Pinheiro GHD, et al. Hematological and physiological changes induced by short-term exposure to copper in the freshwater fish, *Prochilodus scrofa*. *Brazilian Journal of Biology* 2002; 62(4a): 621-631.

14. Rao JV, Shilpanjali D, Kavitha P, et al. Toxic effects of profenofos on tissue acetylcholine esterase and gill morphology in a euryhaline fish, *Oreochromis mossambicus*. *Archives of Toxicology*. 2003; 77(4): 227-232.
15. Camargo MMP, Martinez CBR. Histopathology of gills kidney and liver of a Neotropical fish cage in an urban stream. *Neotropical Ichthyology*. 2007; 5(3): 327-336
16. Matos P, Fontainhas-Fernandes A, Peixoto F, et al. Biochemical and histological hepatic changes of Nile tilapia *Oreochromis niloticus* exposed to carbaryl. *Pesticide Biochemistry and Physiology*. 2007; 89(1): 73-80.
17. Velcheva I, Arnaudov A, Georgieva E. Influence of zinc on gill morphology of Gibelio carp (*Carasius*). *Ecologia Balkanica*. 2010a; 2: 19-23.
18. Velcheva I, Tomova E, Arnaudova D, et al. Morphological investigation on gills and liver of freshwater fish from dam lake "Studen Kladenets". *Bulgarian Journal of Agricultural Science*. 2010b ; 16: 364-368.
19. Sousa DBP, Almeida SZ, Carvalho-Neta RNF. Histology biomarkers in two estuarine cat fish species the Maranhense Coast. *Arquivo Brasileiro de Medicina Veterinaria e Zootecnia* 2013a; 65(2): 369-376.
20. Sousa DBP, Almeida ZS, Carvalho-Neta RNF. Integrated analysis of two biomarkers in *Sciades zbergii* (Arlidae, Siluriformes), to assess the environmental impact at Sao Marcos ? *Latin American Journal of Aquatic Research*. 2013; 45: 305-312.
21. Jha JK, Ranjana, Kumar P, Mishra AP-Histopathological Changes in the gills of *Channa gachua*, an air breathing teleost after short term exposure of hostathion. *Bioscan* 29(3) : 925-929.
22. Velmurugan B, Selvanayagam M, CengizEI, Unlu E- Histopathological changes in the gill and liver tissues of freshwater fish, *Cirrhinus mrigala* exposed to dichlorvos. *Braz. Arch. Biol. Technol*. 2009; 52(5): 1291-1296.
23. Prashanth MS, Sayeswara HA, Goudar MA- Effect of sodium cyanide on behavior and respiratory surveillance in freshwater fish *Labeo rohita* (Ham). *Recent Research in Science and Tech*. 2011; 3:24-30.
24. Kumar SV, Pascal LF, Tennyson S, Pandeeswari M, Dhinamals K, Persis D, Raveen R, Arivoli S, Meeran M- Histopathological studies of *Anabas testudineus* Bloch 1792 on exposure to aquatic toxicants of Buckingham canal, Chennai, Tamil Nadu, India. *International J. Biology Res* 3(2): 125-133.
25. Parashar RS, Banerjee TK- Toxic impact of lethal concentration of lead nitrate on the gills of air breathing catfish *Heteropneustes fossilis* (Bloch). *Vet Arhiv*. 2002; 72: 167-183.